

5

**Abdominal Hernia in a Common Mynah (*Acridotheres tristis*) Associated with Hepatic Lipidosis and Concurrent with Respiratory Aspergillosis**

10

**Running title:** Hepatic lipidosis, hernia & aspergillosis in a mynah

Moein Khodayari<sup>1</sup>, Amir Asghari Baghkheirati<sup>1</sup>, Seyed Mostafa Peighambari<sup>1</sup>, Sara Shokrpour<sup>2</sup>

15

**Jamshid Razmyar<sup>1\*</sup>**

<sup>1</sup> Department of Avian Diseases, Faculty of Veterinary Medicine, University of Tehran, Tehran, Iran.

<sup>2</sup> Department of Pathology, Faculty of Veterinary Medicine, University of Tehran, Tehran, Iran.

20

**Abstract**

**Background:** Hepatic lipidosis and hernia, are problems in pet birds that can occur due to different causes. Aspergillosis, is also an important disease of pet birds.

**Objectives:** This article aims to report an interesting case which by our knowledge is the first one in Iran.

25 **Methods:** A 7-year-old male common mynah was presented to the clinic of avian diseases, the University of Tehran with a history of severe dyspnea, anorexia, feeding with a high energy diet, and travel to the northern parts of Iran in its history. Unfortunately, the bird died. Necropsy was done to diagnose the cause of death. Tissue samples from abdominal viscera, lungs, and air sacs were collected and fixed in 10% buffered formalin and stained with hematoxylin and eosin method. Bacterial cultures from the liver and bone marrow were performed on Blood and MacConkey agars.

30 **Results:** In carcass external examination, abdominal swelling, hernia, and yellow discoloration under the skin were obvious. At necropsy, the liver was enlarged with yellowish discoloration. There was also a high amount of fat around the abdominal viscera. Microscopically, fatty change, hepatocellular necrosis, and hemorrhages were seen in the liver. There were also vegetative forms of *Aspergillus* on the lung's parenchyma and air sacs. No bacterial growth was observed.

35 **Conclusions:** Pet birds are usually restricted to a cage by minimal contact with other birds, thus diseases related to management are common among these birds, so keeping in suitable conditions with an appropriate balanced diet is important for their health.

**KEYWORDS:** Abdominal hernia, Aspergillosis, Common mynah, Hepatic lipidosis, Necropsy

40

#### Case history

A 7-year-old male common mynah was presented to the clinic of avian diseases, the University of Tehran with severe signs of dyspnea. The owner had not noticed the onset of dyspnea and his main complaint was enlargement in the abdominal area. Based on the owner's information, the bird was fed

45

with a diet of high energy (Broiler Finisher diet, Pars Animal and Poultry Feed Ltd, Iran). There was also a history of travel to the northern parts of Iran 6 days before the presentation. The bird was off-fed for 2 days and droppings were watery green. There was also no history of drug use by the owner.

### **Clinical presentation**

50           The bird was perch-off and had severe dyspnea with open mouth breathing and tail bobbing. The wings were dropped and there were obvious hernia and enlargement in the abdominal area (Figure 1.a). The yellow discoloration was present beneath the skin of the area which was an obstacle to the observation of abdominal viscera. Radiography and ultrasonography were proposed to the owner as an aid for diagnosis. Due to the emergency condition of the bird, we first decided to nebulize with  
55   Aminophylline, but before any intervention, the bird died after seizure-like movement. After death, the bird was weighed and its weight was 126 grams.

The cage of the bird was also unsuitable. It was small and the feeder had a lot of feed. The cage floor was dirty and unsanitary and the perch was made of plastic.

### **Diagnostic Testing**

60           Necropsy was done to diagnose the cause of death. The liver was enlarged with round borders and yellow discoloration (Figure1.b). There were copious amounts of fat around the intestine, proventriculus, gizzard, some parts of the lungs and kidneys. (Figure1.c)

Tissue samples of liver, duodenum, proventriculus, gizzard, lung, air sacs, and kidneys were taken for histopathology. The samples were fixed in 10% neutral buffered formalin and routinely  
65   processed, dehydrated, and embedded in paraffin wax, sectioned at 5 µm in thickness, and stained with hematoxylin and eosin method.

Microscopically, fatty change, hepatocellular necrosis, and hemorrhages were seen in the liver. The fatty change was characterized by numerous, fine to large intracytoplasmic vacuoles distorting the hepatic cords. Fatty changes were also observed in the kidneys. A massive development of the vegetative forms of the fungus, containing a large number of conidiophores, hyphae, and spores were observed in the lung's parenchyma and air sacs in H&E staining (Figure 2.a & b).

Cultures from the liver and bone marrow were also performed on Blood and MacConkey agar, but there was no bacterial growth.

### **Assessment**

Hepatic lipidosis is a metabolic disorder occurring in a wide variety of avian species (Schmidt et al., 2015). It is not a specific disease entity but can occur due to disturbance in normal lipid metabolism (Hochleithner et al., 2006). In this condition, lipids accumulate in the liver. It is common in both adult and hand-fed neonates (Hochleithner et al., 2006). There are different etiologies for hepatic lipidosis in adult birds (Beaufre and Taylor, 2013; Vali et al., 2020; Hochleithner et al., 2006). High fat-low protein diet, thyroid dysfunction, and restricted exercise are some of them (Beaufre and Taylor, 2013). Increased lipogenesis which can be a sequel of estrogen activity, diabetes mellitus, stress, and estrogen-like activity of pesticides are other factors (Beaufre and Taylor, 2013; Hochleithner et al., 2006). Nutritional deficiencies, as essential fatty acids, sulfur amino acids and vitamins like biotin, B1, B6, and B12 can also be a probable cause (Beaufre and Taylor, 2013; Hochleithner et al., 2006). Acute release of fatty acids from adipose tissues may have a role in this disorder (Beaufre and Taylor, 2013; Vali et al., 2020). Some references have named mycotoxins and drugs (which impair secretion of lipoprotein) as a probable cause (Hochleithner et al., 2006).

In chicks, the most common cause is overfeeding with high energy formula (Beaufre and Taylor, 2013). It was claimed that in neonates, high levels of avidin in yolk sac remnants may reduce

90 available biotin, which can lead to reduced hepatic gluconeogenesis (Davies, 2000). Based on published information, some species of parrots (like Amazon parrots, galah cockatoos, budgerigar, and lorikeets) are more susceptible to hepatic lipidosis (Beaufreere and Taylor, 2013).

History and clinical signs, clinical pathology, diagnostic imaging, and hepatic biopsy are some of the methods for diagnosis in live birds. Post mortem gross lesions in the liver and sampling for  
95 histopathology can lead to definitive diagnosis (Grunkemeyer, 2010).

In general, treatment includes cool oxygenation in cases of dyspnea, non-lactated fluid therapy to rehydrate the bird and detoxify the body, treatment of secondary infections, nutritional balance, using metabolic aids like silymarin and preparing conditions for body exercise (Hochleithner et al., 2006).

The etiology of abdominal hernias in birds is unknown, but in general, they can be congenital and  
100 acquired (Bennett and Harrison, 1994; Anderson et al., 2018). Some studies consider them as a separation in the aponeurosis of abdominal musculature (Bennett and Harrison, 1994; Macwhirter, 1994). Other studies have proposed that due to the absence of opening in muscular aponeurosis, they are not a true hernia (Anderson et al., 2018).

Some predisposing factors have been described to the hernia such as hyperestrogenism, lack of  
105 exercise, malnutrition, trauma, and egg-laying behavior. (Bennett and Harrison, 1994; Macwhirter, 1994; Amer et al., 2018) Besides, some hernias were associated with hepatic lipidosis, reproductive tract problems (like egg yolk peritonitis), peritoneal cyst (Bennett and Harrison, 1994; Macwhirter, 1994), and intraabdominal lipoma (Razmyar et al., 2005). Anyway, if the hernia becomes larger, it can be a cause of skin ulcers and hemorrhage (Bennett and Harrison, 1994; Macwhirter, 1994; Amer et al., 2018).

110 Treatment may not be the same in different cases. In some cases, especially when trauma is present, surgical repair is recommended (Murphy et al., 2018). In some cases, like hepatic lipidosis,

treatment can reduce organ size which may lead to the elimination of hernia without surgery (Macwhirter, 1994; Barboza et al., 2018).

115 Aspergillosis is a fungal, noncontagious disease of both wild and caged birds (Nardoni et al., 2006; Carrasco and Forbes, 2016; Melo et al., 2020; Leishangthem et al). Infection is usually by inhalation of conidia or spores of pathogenic species of *Gnus Aspergillus*, especially *A. fumigatus*. (Nardoni et al., 2006; Carrasco and Forbes, 2016). They are ubiquitous and all birds are susceptible to infection (Nardoni et al., 2006). The source of infection is usually contaminated feed, litter, and soil (Carrasco and Forbes, 2016). The disease is primarily restricted to the respiratory system but can be systemic and involve other  
120 organs (Nardoni et al., 2006; Carrasco and Forbes, 2016).

There are some characteristics in avian species that predispose them to the disease. For example unique anatomy of the respiratory tract and higher body temperature accelerates fungal growth. Other factors include vitamin deficiency, especially vitamin A, chronic stress, overcrowding, unsanitary condition, malnutrition, treatment with corticosteroids, and respiratory irritants (Nardoni et al., 2006; Leishangthem et al., 2015; Krautwaldjunghanns et al., 2015).  
125

The disease can be acute or chronic. The acute form is more common in chicks and results from inhaling a high number of spores. It can develop in less than a week. Clinical signs of this form are not specific and can be confusing. Dyspnea, cyanosis, anorexia, polydipsia, and fetid diarrhea are some of the probable clinical signs (Leishangthem et al., 2015; Fischer and Lierz, 2015).

130 The chronic form is more common in older birds and takes weeks or months to develop. Clinical signs may vary depending on the site of infection. Different references have reported different clinical signs in the respiratory system, skin, eyes, GI tract, nervous system, bones, joints, and abdominal viscera (Leishangthem et al., 2015; Fischer and Lierz, 2015).

Diagnosis of the disease in the live birds is very difficult, especially in acute cases (Leishangthem et al., 2015). Some diagnostic tests like biochemistry, hematology, endoscopy, and imaging can be helpful (Fischer and Lierz, 2015; Savelieff et al., 2018). Definitive diagnosis is based on histopathology from post mortem samples or biopsy (Elad and Segal, 2018; Leishangthem et al., 2015). Fungal culture from suspected organs can also be helpful (Savelieff et al., 2018). Other diagnostic tools like ELISA and agar gel immunodiffusion are not common and never should be used alone (Fischer and Lierz, 2015). Treatment by antifungal drugs is complex and depends on the site and extent of infection (Leishangthem et al., 2015).

In Iran, keeping birds as a pet (mainly from psittacines and passerines) is increasing in recent years. All domestic birds have not the same nutritional and management requirements. Unfortunately, most of the owners are not familiar with these needs. Thus, malnutrition and mismanagement are usual findings in the most cases presented to avian clinics.

In this case, according to the history and clinical signs, it seems that hepatic lipidosis is a result of feeding with a high energy broiler diet and less activity due to the small size of the cage. Feeding mynah with broiler pellet is very usual in Iran because it is similar to mynah's pelleted feed. There is another report of hepatic lipidosis associated with high energy feed in Iran (Madani et al., 2012).

As described earlier, hepatic lipidosis can make birds prone to hernias. There is a report in a red lory that supports their association (Langlois and Jones, 2001). In this case, the most probable cause of hernia is hepatic lipidosis. This condition causes enlargement of the liver which puts the respiratory tract under pressure. This pressure can lead to lower respiratory capacity.

Damp climate, which is common in northern parts of Iran in summer, encourages fungal growth. The acute form of aspergillosis in birds is a result of inhaling a high amount of spores (Leishangthem et al., 2015). Because the bird had traveled to the north part of Iran, it is possible that had exposure to high

numbers of fungal spores which may lead to acute aspergillosis. This disease has exacerbated the effect of hepatic enlargement on the respiratory tract. This can elucidate the cause of death and premortem clinical signs.

160           There is no published report of simultaneous hepatic lipidosis, hernia, and aspergillosis in any domestic bird in Iran. Because avian medicine is a relatively new field in Iran and the world, so reporting these kind of cases could be helpful for developing this field (Pollock et al., 2016). It can also help avian veterinarians and owners to promote their knowledge in captive animals management and potential diseases of domestic birds which are rare in the wild bird populations.

165

## References

1. Amer, M. S., Hassan, E. A., Torad, F. A. (2018). Radiographic and ultrasonographic characteristics of ventral abdominal hernia in pigeons (*Columba livia*). J Vet Med Sci., 80(2), 292–296. <https://doi.org/10.1292/jvms.17-0517>. PMID: [29237997](https://pubmed.ncbi.nlm.nih.gov/29237997/)
- 170 2. Anderson, K., Brandão, J., Mans, C. (2018). Lateral body wall herniation involving the oviduct in two psittacine birds. J Avian Med Surg., 32(4), 328-335. <https://doi.org/10.1647/2017-320>. PMID: 31112647
3. Barboza, T. K., Beaufrère, H., Chalmers, H. (2018). True coelomic hernia and herniorrhaphy in a yellow-crowned amazon parrot (*Amazona ochrocephala*). J Avian Med Surg., 32(3), 221-225. <https://doi.org/10.1647/2017-321>. PMID: **30204013**
- 175 4. Beaufrere, H., Taylor, W. M. (2013). Hepatic lipidosis. In: Clinical Veterinary Advisor: Birds and Exotic Pets. Mayer, J., Donnelly, T. (eds.). Elsevier Saunders. St. Louis, USA. P. 194-196.



5. Bennett, R. A., Harrison, G. J. (1994). Soft tissue surgery. In: Avian Medicine: Principles and Application. Ritchie, B. W., Harrison, G. J., Harrison, L. R. (eds.). Wingers Publishing. Florida, USA. p. 1097–1136.
- 180 6. Carrasco, D. C., Forbes, N. A. (2016). Aspergillosis: update on causes, diagnosis and treatment. Companion Anim., 21(1), 50-57. <https://doi.org/10.12968/coan.2016.21.1.50>.
7. Davies, R. R. (2000). Avian liver diseases: Etiology and pathogenesis. J Exot Pet Med., 9(3), 115-125. <https://doi.org/10.1053/ax.2000.7138>.
8. Elad, D., Segal, E. (2018). Diagnostic aspects of veterinary and human aspergillosis. Front Microbiol.,  
185 9, 1303. <https://doi.org/10.3389/fmicb.2018.01303>. PMID: 29977229.
9. Fischer, D., Lierz, M. (2015). Diagnostic procedures and available techniques for the diagnosis of aspergillosis in birds. J Exot Pet Med., 24(3), 283-295. <https://doi.org/10.1053/j.jepm.2015.06.016>.
10. Grunkemeyer, V. L. (2010). Advanced diagnostic approaches and current management of avian hepatic disorders. Vet Clin North Am Exot Anim Pract., 13(3), 413-427.  
190 <https://doi.org/10.1016/j.cvex.2010.05.005>. PMID: 20682427.
11. Hochleithner, M., Hochleithner, C., Harrison, L. D. (2006). Evaluating and treating the liver. In: Clinical Avian Medicine. Harrison, G. J., Lightfoot, T. L. (eds.). Volume I. Spix Publishing. Palm Beach, USA. p. 441-449.

12. Krautwaldjunghanns, M. E., Vorbrüggen, S., Böhme, J. (2015). Aspergillosis in birds: an overview of  
195 treatment options and regimens. *J Exot Pet Med.*, 24(3), 296-307.  
<https://doi.org/10.1053/j.jepm.2015.06.012>.
13. Langlois, I., Jones, M. P. (2001). Ventral abdominal hernia associated with hepatic lipidosis in a red  
lori (*Eos bornea*). *J Avian Med Surg.*, 15(3), 216-222. [https://doi.org/10.1647/1082-  
6742\(2001\)015\[0216:VAHAWH\]2.0.CO;2](https://doi.org/10.1647/1082-6742(2001)015[0216:VAHAWH]2.0.CO;2).
- 200 14. Leishangthem, G. D., Singh, N. D., Brar, R. S., Banga, H. S. (2015). Aspergillosis in avian species: A  
review. *J Poult Sci Technol.*, 3(1), 01-14.
15. Macwhirter, P. (1994). A review of 60 cases of abdominal hernias in birds. *Proc Annu Conf Assoc  
Avian Vet.*, 27-37.
16. Madani, S. A., Hatamkhani, A., Soroori, S. (2012). Hepatic lipidosis in a common mynah  
205 (*Acridotheres tristis*) associated with pododermatitis and consumption of broiler pelleted feed. *Iran J Vet  
Med.*, 6(4), 279-283. [10.22059/IJVM.2012.30226](https://doi.org/10.22059/IJVM.2012.30226).
17. Melo, A. M., da Silva-Filho, R. P., Poester, V. R., von Groll, A., Fernandes, C. G., Stevens, D. A.,  
Sabino, R., Xavier, M. O. (2020). Aspergillosis in free-ranging aquatic birds. *Med Mycol Case Rep.*,  
<https://doi.org/10.1016/j.mmcr.2020.04.005>. PMID: 32405453
- 210 18. Murphy, B., Fitzgerald, B., Olsen, G., Speer, B. (2018). Diagnosis and Surgical Repair of an Acute  
Abdominal Wall Hernia and Partial Cloacal Strangulation in a Yellow-Naped Amazon Parrot (*Amazona  
auropalliata*). *J Avian Med Surg.*, 32(4), 336-341. <https://doi.org/10.1647/2017-330>. PMID: 31112648.
19. Nardoni, S., Ceccherelli, R., Rossi, G., Mancianti, F. (2006). Aspergillosis in *Larus cachinnans  
micaellis*: survey of eight cases. *Mycopathologia.*, 161(5), 317-321. [10.1007/s11046-006-0012-2](https://doi.org/10.1007/s11046-006-0012-2). PMID:  
215 16649081.

20. Pollock, C., Klaphake, E., Wellehan Jr, J.F.X. (2016). Avian medicine: an overview. In: Current Therapy in Avian Medicine and Surgery. Speer, B. L. (ed). (1st ed). Elsevier Saunders. California, USA. p.1-8.
21. Razmyar, J., Vajhi, A., Peighambari, S., Dezfoulian, O. (2005). Case Report: Lipoma in a budgerigar  
220 (*Melopsittacus undulatus*). **J Fac Vet Med Univ Tehran.**, 297-298.
22. Savelieff, M. G., Pappalardo, L., Azmanis, P. (2018). The current status of avian aspergillosis diagnoses: Veterinary practice to novel research avenues. *Vet Clin Pathol.*, 47(3), 342-362. <https://doi.org/10.1111/vcp.12644>. PMID: 30117564
23. Schmidt, R. E., Reavill, D. R., Phalen, D. N. (2015). Liver. In: Pathology of Pet and Aviary Birds.  
225 (2nd Ed). Wiley – Blackwell. Hoboken, New Jersey, USA. p. 95-125.
24. Vali, Y., Molazem, M., Soroori, S., Madani, S. A., Hassanzadeh, M. (2020). Clinical Application of Three-Dimensional Quantitative Ultrasonography in Presumptive Diagnosis of Hepatic Lipidosis in Common Mynahs (*Acridotheres tristis*). *Res Sq.* <https://doi.org/10.21203/rs.3.rs-20199/v1>.

230

فتق شکمی در یک مینای معمولی مرتبط با لیپیدوز کبدی و همزمانی با آسپرژیلوز تنفسی

معین خدایاری<sup>1</sup>، امیراصغری باغخیراتی<sup>1</sup>، سید مصطفی پیغمبری<sup>1</sup>، سارا شکر پور<sup>2</sup>، جمشید رزم یار<sup>1\*</sup>

1) گروه بیماری‌های طیور، دانشکده دامپزشکی، دانشگاه تهران، تهران، ایران.

235

2) گروه پاتولوژی، دانشکده دامپزشکی، دانشگاه تهران، تهران، ایران.

زمینه مطالعه: لیپیدوز کبدی و فتق مشکلاتی در پرندگان خانگی هستند که می‌توانند به دلایل مختلفی ایجاد شوند. همچنین، آسپرزیلوز یک بیماری مهم در پرندگان خانگی است.

240

هدف: هدف این مقاله گزارش یک کیس جالب است که با توجه به اطلاعات ما اولین مورد در ایران است.

روش کار: یک مینای معمولی 7 ساله نر با تاریخچه‌ای از سختی تنفس شدید، بی‌اشتهایی، تغذیه با جیره پرانرژی و سفر به مناطق شمالی ایران در تاریخچه‌ی خود، به کلینیک بیماری‌های پرندگان، دانشگاه تهران ارجاع شد. متاسفانه، پرنده تلف شد. کالبدگشایی به منظور تشخیص علت مرگ انجام گرفت. نمونه‌های بافتی از احشای محوطه بطنی، ریه‌ها و کیسه‌های هوایی جمع‌آوری شده و در بافر فرمالین 10% فیکس شده و به روش هماتوکسیلین و انوزین رنگ‌آمیزی شدند. کشت‌های باکتریایی از کبد و مغز استخوان در Blood agar و MacConkey agar انجام گرفتند.

245

نتایج: در معاینه بیرونی لاشه، تورم ناحیه شکم، فتق و تغییر رنگ زرد در زیر پوست مشهود بود. در کالبدگشایی، کبد متورم و همراه با تغییر رنگ مایل به زرد بود. همچنین مقادیر زیادی چربی در اطراف احشای شکمی وجود داشت. با استفاده از میکروسکوپ، در کبد، تغییر چربی، نکروز سلول‌های کبدی و خون‌ریزی مشاهده شدند. همچنین اشکال رویشی آسپرزیلوس در پارانشیم ریه و کیسه‌های هوایی وجود داشتند. هیچ رشد باکتریایی مشاهده نشد.

250

نتیجه‌گیری نهایی: معمولاً پرندگان خانگی در قفس محصور بوده و تماس کمی با سایر پرنده‌ها دارند، بنابراین بیماری‌های مرتبط با مدیریت در بین این پرنده‌ها رایج است، بنابراین نگهداری در شرایط مناسب به همراه جیره متعادل و مناسب برای سلامت آن‌ها مهم است.

واژه‌های کلیدی: فتق شکمی، آسپرزیلوز، مینای معمولی، لیپیدوز کبدی، کالبدگشایی

255

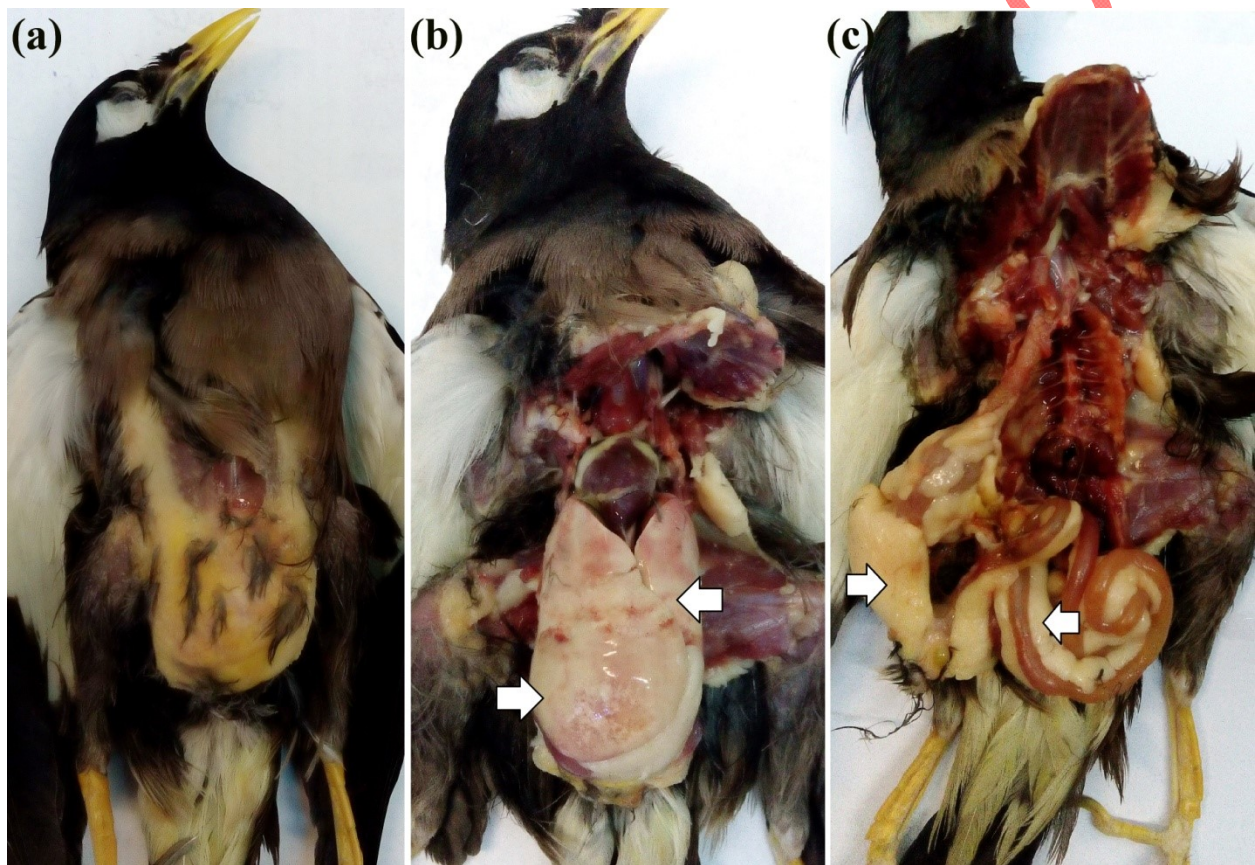
## Figure Captions

**Figure 1:** carcass external examination & necropsy findings

a. Abdominal enlargement and hernia

260 b. Enlarged yellowish liver with rounded borders

c. Copious amount of fat around abdominal viscera



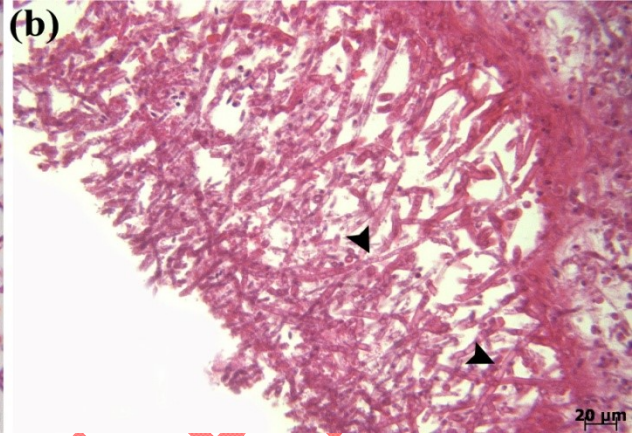
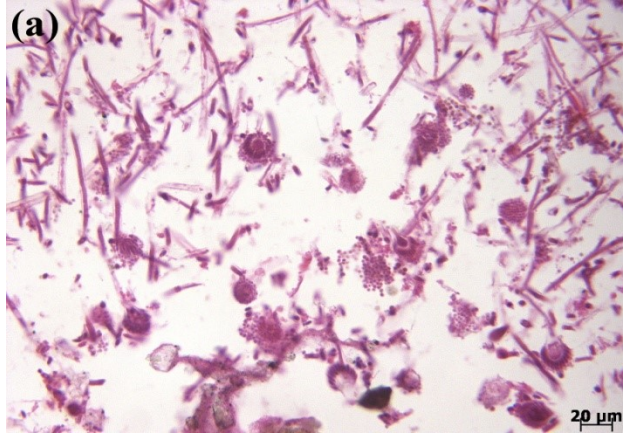
270

**Figure 2:**

a. Fungal conidiophores in air sac section stained with H & E method (x 40)

b. Fungal hyphae in air sac section stained with H& E method (x 40)

275



280

285

Uncorrected