RESEARCH PAPER



# Assessment of Environmental Parameters in Pre- and Post-Monsoons at Chabahar Bay, Gulf of Oman

Agah, H.<sup>1\*</sup>, Saleh, A.<sup>1</sup> and Jalili, M.<sup>2</sup>

<sup>1</sup> Assistant Professor, Iranian National Institute for Oceanography and Atmospheric Science, Tehran, Iran.

<sup>2</sup> M.Sc., Iranian National Institute for Oceanography and Atmospheric Science, Tehran, Iran.

© University of Tehran 2021

Received: 26 Oct. 2019;

Revised: 13 Sep. 2020;

Accepted: 19 Sep. 2020

ABSTRACT: This study is the first investigation to assess the variations of physical and chemical characteristics and biodiversity of planktons in offshore water column and chlorophyll-a and b, during the two monsoons at Chabahar Bay, to evaluate the water quality. To this end, 27 surface water samples in pre-monsoon (May, 2012) and totally 60 surface and deep water samples in post-monsoon (December, 2012) were collected from 9 and 10 stations at depths between 3.8 to 13.6 m in Chabahar Bay, respectively. The results showed that water salinity and pH with low variations were relatively higher in post-monsoon. The average of water alkalinity levels in pre- (2.42±0.02 mmol H<sup>+</sup>/kg) and post- (2.44±0.01 mmol/kg) monsoons were comparable to that of oceanic surface water (2-2.5 mmol H<sup>+</sup>/kg). In this study, 66 phytoplankton genus and species belonging to 13 groups were identified in pre-monsoon. Results demonstrated that nutrients were at higher levels inside the Chabahar Bay. Moreover, the physicochemical parameters of water samples were investigated and compared with international standards and data from other marine ecosystems. The results indicated that the water quality falls within the stipulated range of acceptability and sampling area can be classified as a good, stable, and healthy aquatic ecosystem.

Keywords: Chabahar Bay, Gulf of Oman, Nutrients, Phytoplankton, Water Quality.

### **1. Introduction**

It is well known that seawater resources are under threat by human activities (Salvi et al., 2014). Water quality in marine environment plays a very important role in human wellbeing, and has a prominent impact on metabolic pathways in marine organisms (Mearns et al., 2013). Therefore, minor variations in some parameters such as pH and nutrients might play key roles in growth and health of biota. Marine phytoplankton which communities contribute to primary productivity are largely dependent on nutrient availability, light penetration and quality of water column in coastal areas (Basu et al., 2018). Among various nutrients, nitrogen, phosphorous and silicon are considered to be more important than the others, due to being crucial for phytoplankton abundance, growth and metabolism (Shanthi et al., 2014). The Gulf of Oman is influenced by

<sup>\*</sup> Corresponding author E-mail: aaagah\_hom@yahoo.com

upwelling events that bring a great amount of nutrients into the upper ocean, hence enhancing primary productivity and ultimately the fisheries.

Physicochemical characteristics are indeed vital water quality parameters for monitoring due to their instability (Efe et al., 2005), since significant variations in physicochemical parameters affect the quality of water resources. Therefore, it would be impotent to monitor water quality of all marine sources.

In this study, total alkalinity of seawater, which is one of the required parameters to evaluate the carbonate chemistry of seawater, was investigated. The water quality of Chabahar Bay on the Makran coastline in Sistan and Baluchestan Province, Southeast of Iran, was studied. Chabahar Bay is a free port, industrial zone and an important shipping route for the oilproducing countries in Persian Gulf.

Little studies have focused on physicochemical data in Chabahar Bay, and hence it seems required to continue and perform further investigations to protect ecosystem. this valuable The main objectives of the present study were: 1) To determine water quality parameters (physicochemical characteristics, nutrients, alkalinity and chlorophylls) from the Chabahr Bay, Oman Sea; 2) To investigate the abundance and biodiversity of planktons in water column; 3) To discover the relationships between the abundance of phytoplankton and water physicochemical parameters and, 4) To compare the results from different studied stations with each other and with data from other marine ecosystems.

## 2. Materials and Methods

In recent years, Chabahar Bay has become more important, due to the increase in human population, urbanization and accelerated developmental activities. The special importance of Chabahar Bay is its  $\Omega$ shape and the limited water circulation. The study area has humid climate, hot summers and moderate winters with maximum and average depths of 22 and 12 meter, respectively. Fishing and marine commerce are the main activities in the study area (Fazel, 2008).

We utilized glass/ reference electrode (Hach, IntelliCAL PHC 101), calibrated by NBS buffers (accuracy of +0.02, precision of +0.001) to measure surface water temperature and pH. Salinity was determined using a conductivity probe (Hach, IntelliCAL CDC401 with the precision of +0.1), calibrated by a certified seawater. Dissolved oxygen was measured using a calibrated dissolved oxygen optical sensor (HACH, IntelliCAL LDO101 luminescent/optical dissolved oxygen probe). In addition, CTD was used to obtain the physicochemical parameters of surface and deep water (temperature, salinity) at pre- and post-monsoons. Geographical characteristics of sampling locations were recorded by GPS (Geographic Positioning System). All sensors of the CTD were calibrated according to the guidelines of the related manufacturers, based on 1 sec. per data. In each station, CTD was sank 1 m/min to prepare water profile parameters (temperature, pH, turbidity, salinity, and dissolved oxygen). All data of the CTD was controlled and initially processed in excel to detect outliers. The clarity of water samples was measured using Secchi disc.

In order to detect the concentrations of different nutrients (nitrite. nitrate. phosphate and silicate), water organic carbon, alkalinity, phytoplankton species and chlorophyll-a and b, surface water samples were collected in pre- and postmonsoons from geographical situations located at 25° 15' 33" to 25° 26' 3" of latitudey, and 60° 15' 21" to 60° 45' 18" of longitude, in Chabahar Bay (Table 1). The sampling locations were: 1) Tiss; 2) Konarak; 3) Desalination plant; 4) Entrance of Chabahar Bay; 5) Posm and; 6) Ramin (Figure 1). The selected sampling sites represented the most important harbors, desalination plants, and industrial regions in the area.

Totally 27 surface water samples in premonsoon (May, 2012), 30 surface and 30 deep water samples in post-monsoon (December, 2012) were collected from 9 and 10 stations at depths between 3.8 to 13.6 m in Chabahar Bay, respectively (Figure 1). Jellyfish bloom at deeper parts of Ramin station in pre-monsoon limited the sampling and measurements of the physicochemical parameters at this station.

Triplicate surface water samples (collected from a depth of 1 m) in premonsoon as well as the surface and bottom water samples in post-monsoon were collected using Niskin bottle (2 L).

In order to determine the concentrations of the nutrients, an aliquot of each water sample (100 mL) was filtered using 0.45-µ cellulose acetate filter, collected in labeled polyethylene bottles, carried to the laboratory on ice, and analyzed during 24 h. Nitrite was detected based on pink diazo dye formation and its colorimetry. Nitrate was measured after its reduction to nitrite using cadmium powder. Phosphate was based detected on colorimetric phosphomolybdate. determination by Dissolved nutrients such as nitrate, nitrite, phosphate and silicate contents were measured using HACH spectrophotometer (Ray Light UV 9200, MOOPAM, and ROPME, 1999).

Water samples for determination of total alkalinity were collected in 500-mL glass bottles, and 100 µL of a saturated mercury chloride (HgCl<sub>2</sub>) solution (II)was subsequently added to stop the biological activities (Dickson et al., 2007). Samples were stored at 4 °C until laboratory analysis. Total alkalinity was determined by potentiometric open-cell titration (Dickson et al., 2007, SOP03b) using a digital 715 Dosimat titrator (Hydro-bios). For pH measurements, a Sartorius pH meter (with precision of  $\pm 0.01$ ) was used.

Total organic carbon in water samples was analyzed using TOC Analyzer (SGE, ANATOC Seri II Australia; Agah et al., 2014). Detection limit of TOC analyzer was 50 ppb. The concentrations of TOC in all samples were higher than detection limit. The recovery of methodology was 97±2%.

In order to determine the concentrations of chlorophylls, one liter of post-monsoon surface and bottom water samples were filtered through 47 mm glass fiber filters. According to the Standard Method (10200 H), chlorophyll-a and b were extracted and measured by HPLC using UV (absorption wavelength of 440 nm) and fluorescence (excitation and emission wavelengths of 430 and 600 nm) detectors, respectively.

Surface (n = 27 and 30) and sea bottom water samples (n = 30, ~1 m depth from bed) in pre- and post-monsoon sampling periods were transferred to dark bottles, preserved in Lugol's iodine, followed by storage. Identification of phytoplanktons and their abundance was performed using an inverted microscope (Axiostar S100, Ziess) with 40X objectives (Sedgwick-Rafter Method) and various taxonomic keys (Al-Kandari et al., 2009; Xiang et al., 2019; ROPME, 2012, Karlson, 2010; AlgaeBase).

Results were reported as the average of triplicate analyses and their corresponding standard deviations. Statistical analysis was performed using SPSS (version 2017) software. The Pearson correlation matrix was carried out using SPSS statistical software (Version 16.0 for Windows). A Kolmogorov-Smirnov test was performed normality evaluate the of data to distribution. In addition, one-way ANOVA was used to assess significant differences between the mean levels of the elements at different sampling stations. A P-value equal or lower than 5% indicated that significant relationship between the corresponding variables exists.

The Mann-Whitney analysis was used to evaluate the significant differences in phytoplankton density at the studied stations using SPSS statistics, version 21 software.

## 3. Results and Discussion

Water quality of Chabahar Bay, the main Iranian omega shape gulf in the Makran zone, Gulf of Oman with high transport and trade potential, has been affected by several factors in last decades, as a result of anthropogenic activities (Burt et al., 2016; Agah, 2018). Hence, studying the water physicochemical characteristics of the gulf can guide us to protect this ecosystem during its developing programs.

Temperature (or its changes) as a basic property of water is important in regulation of many physiological processes in marine organisms and is therefore one of the most important water quality attributes in aquaculture (IEPA, 2001); it controls water metabolism and specifies the habitat area for aquatic life (Ding and Elmore, 2015).

Owing to the high level of natural variability in pH, salinity and DO in coastal marine waters both temporally and geographically, it is not possible to provide fixed guideline values for temperature. The temperature values of the surface water (Figure 2) at all stations were comparable throughout the study period, varying from 26 (Posm) to 28.6 (Tiss) °C and from 24.3 (Konarak) to 26.4 (Ramin) °C in pre- and post-monsoons, with average of 26.6 and 25.5 °C, respectively.

Variation of surface water temperature in pre- and post-monsoons inside Chabahar Bay were relatively higher than that of other stations, which could be related to less water exchange from inside the semi-closed Average of deep layer water bay. temperature was 25.1 °C in post-monsoon. According to NOAA Coral Reef Watch (2019), the annual minimum and maximum sea surface temperature variation in Chabahar Bay at 2017 was belonged to February (22.8 °C) and June (30.3 °C) with average value of 27.5 °C. Results revealed that sea surface temperature in all the studied stations was within the range of the area.

Salinity, pH and temperature of the surface water were uniform at all the stations and exhibited only a narrow variation. The range and average values of pH and salinity of surface water in premonsoon were 8.03 (Posm)-8.15 (Desalination plant) ( $8.12\pm0.04$ ) and 36 (Posm)-37.3 (Tiss) ( $36.6\pm0.37$ ) ‰, and in post-monsoon were 8.13 (Posm)-8.21 (Ramin) ( $8.18\pm0.04$ ) and 36.3 (Ramin)-37 (Posm) ( $36.7\pm0.36$ ) ‰ (Table 2).

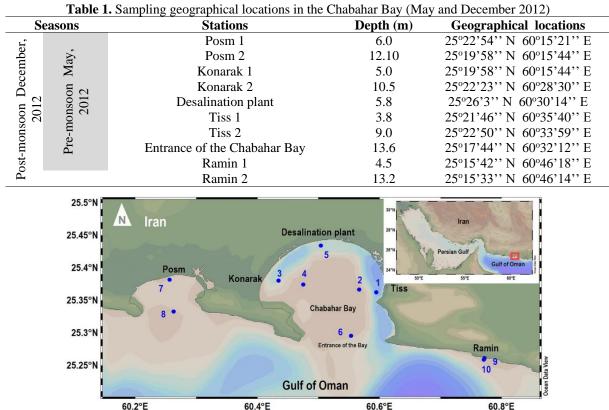


Fig. 1. The location of the sampling site in the Chabahr Bay

#### Agah et al.

	Dej (n		Te	nperat °C	ture		рН		Sa	linity	‰	Do (mg		TOC (ppm)	Secc	arity hi disc m)
Station			Pre	Post	post	Pre	Post	post	Pre	post	post	post	post	Pre		
S	Pre	Pre Post	Surface	Surface	Bottom	Surface	Surface	Bottom	Surface	Surface	Bottom	Surface	Bottom	Surface	Pre	Post
Posm 1	9	4.8	25.9	25.4	25.2	8.03	8.18	8.2	36.3	37	37	7.23	7.46	1.41	$\tilde{\omega}$	2.5
Posm 2	12.1	10.6	25.1	26	25.6	8.1	8.18	8.19	36.0	36.7	36.8	7.06	7.15	1.50	1.3	9
Konarak 1	5	4.6	26	24.3	24.2	8.13	8.18	8.19	36.6	36.9	37	7.33	7.54	2.56	1.5	4
Konarak 2	10.5	10	26.1	24.8	24.1	8.13	8.18	8.14	36.8	36.8	37	7.54	6.23	1.04	1.2	ę
Desalinat ion plant	5.8	4.7	26.2	25	24.5	8.15	8.2	8.15	36.6	36.7	36.8	7.04	6.9	1.51	1.75	3.5
Tiss 1	3.8	3.1	28.6	24.9	25	8.11	8.17	8.17	36.8	36.7	36.7	6.62	6.61	2.78	0.9	3.1
Tiss 2	6	8.2	28.5	25	24.9	8.11	8.16	8.17	37.3	36.7	36.8	6.73	6.82	1.5	1.2	8.2
Entrance of Chabahar Bay	13.6	14.3	26.5	26.2	25.1	8.14	8.16	8.13	36.7	36.5	36.9	6.9	6.13	1.68	2.7	8.5
Ramin 1	4.5	4.5	26.9	26.6	26.1	8.15	8.2	8.21	36.3	36.3	36.4	6.96	6.84	1.41	3.5	4.5
Ramin 2	ı	13.3	ı	26.4	26.4		8.19	8.2	ı	36.3	36.4	6.82	6.85		4.5	8.5
Average			26.6±1.2	25.5± 0.8	$25.1 \pm 0.8$	$8.1 {\pm} 0.04$	$8.18 \pm 0.01$	$8.18 \pm 0.03$	$36.6\pm 0.4$	36.7±0.2	$36.8 \pm 0.2$	7.0±0.3	$6.8 \pm 0.5$	$1.7 \pm 0.57$	189.4±93	5.18±2.4

**Table 2.** Physico-chemical parameters of water samples in different station in pre and post Monsoon (May and December 2012)

(Pre: Pre Monsoon and Post: Post Monsoon)

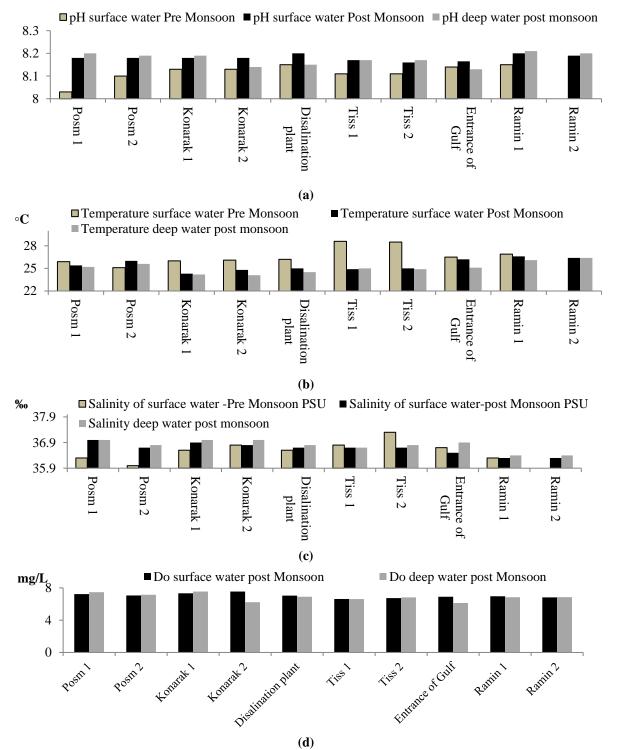


Fig. 2. a) pH; b) Temperature; c) Salinity and; d) Dissolved oxygen of surface and deep layer at pre and post Monsoons in Chabahar Bay (May and December 2012)

Although the salinity of surface waters relatively varied at different stations during two monsoons, their average values (36.6 and 36.7 ‰) were comparable (P > 0.05). In an investigation, Fazeli et al. (2010) assessed seasonal variations of Copepoda in Chabahar Bay. In this study, water sanity was reported to be within the range of 36.7 to 36.9 ‰, which was comparable to that of the present study during the two monsoons. Ibrahim (2010) investigated water quality parameters at southern parts of the Gulf of Oman on February-March 1987, and reported a mean water salinity of 36.70. Comparing the results of this study with those of the present work demonstrated that water salinity values in pre-monsoon were comparable over the last two decades (Ibrahim, 2010).

Average values of pH and salinity for bottom water in post-monsoon were 8.18 and  $36.8\pm0.2$  ‰, respectively. Low variations in pH within the sampling area (Figure 2) during pre- and post-monsoons represented the water homogeneity.

Results demonstrated that the average value of pH in the sampling area was within the range of the surface ocean pH (Raven et al., 2005), producing appropriate conditions for coral reefs (which are very sensitive to any changes in pH) (Omer, 2010).

Obtained results indicated that pH of the surface water in post-monsoon were slightly higher than that at pre-monsoon. Seasonal variations of atmospheric carbon dioxide and phytoplankton activities can influence pH variations in different seasons.

Concentrations of 5.0 mg/L or higher for dissolved oxygen are desirable for fish survival. Low levels of dissolved oxygen are known to be one of the major problems for faunal and floral survival in aquatic environment (Friedrich et al., 2014). When the base DO level is less than the recommended level, that concentration should become the interim guideline at that time (CCME, 2003). The range and average values of dissolved oxygen of surface and deep layer water in post-monsoon varied from 6.6 (Tiss1) to 7.5 (Konarak2)  $(7.0\pm0.3)$  and from 6.13 (Entrance of Chabahar Bay) to 7.54 (Konarak)  $(6.9\pm0.5)$ mg/L, respectively. Hence, the dissolved oxygen levels measured in the present study were considered to be moderate to sustain the aquatic biodiversity. Generally organic wastes and other nutrient inputs from and industrial discharges, sewage agricultural, and urban runoff can result in decreased oxygen levels in some stations (Nasir Khan and Mohammad, 2014).

The pH of an aquatic ecosystem is an important indicator of the water quality. The ideal pH for biological productivity ranges between 6.8 to 8.5 (CCME, 2003; EPA, 1970), while pH values lower than 4

are detrimental to aquatic life (Abowei, 2010) and also allow toxic elements and compounds such as heavy metals to become mobile to aquatic life (APHA, 1995). Waters with pH values from 6.5 to 9.0 would be best for fish production (Olatayo, 2014). The recorded pH values in the present study ranged from 8.1 to 8.28 by a mean value of 8.18 (Table 2), which was within the range of the sea waters (6.5-8.4) (Olatayo, 2014), and was appropriate for aquatic life.

In pre-monsoon, pH value (8.03) and salinity (36.0 ‰) were relatively lower at Posm, as a closed bay (p > 0.05), compared to the other stations (Figure 2), which might be affected by rain fall one day before sampling. This finding could be confirmed by comparison of temperature at different stations.

Figure 3 illustrates the profiles of water temperature and salinity in pre-monsoon as typical measurments by CTD within the studied area (May 2012).

The range and average values of organic carbon at pre-monsoon were 1.04 (Konarak) -2.78 (Coastal parts of Tiss) and  $(1.7\pm0.56)$  ppm, respectively (Table 2).

Surface waters of some stations (Posm, Konarak, Tiss, and Ramin) were analyzed to determine total alkalinity in both pre- and post-monsoons. The averages of surface total alkalinity in pre- and post-monsoons were 2.422  $\pm$  0.02 mmol/Kg and 2.439  $\pm$ 0.01 mmol/Kg (Figure 4), which were within the range of oceanic surface water (2-2.5 mmol/Kg), and slightly higher than data available from GLODAPv2.2020 (Olsen et al., 2020) in the offshore Oman Sea  $(2.409 \pm 0.01)$ , surface water, August 1995). Recently, Saleh et al. (2020) reported total alkalinity of surface waters within the range of 2.413 to 2.425 mmol/Kg for seawater samples from intertidal rocky shores of the Chabahar Bay and Makran Sea (northern Oman Sea) (Saleh et al., 2020).

Results of post-monsoon sampling indicated that total alkalinity of the surface and near-bottom layers in shallower stations (stations of inner Chabahar Bay with depths of less than 5 m) did not show significant differences (P > 0.05), probably due to mixing process forced by tidal currents. Low values of positive correlation coefficients between total alkalinity as a conservative seawater characteristic and salinity ( $r^2 = 0.2$ , p > 0.05), might be related to biological activities (such as biocalcification and photosynthesis), and fresh water input (surface seasonal floods and groundwater inflows) (Xue and Cai, 2020).

Nutrient input often leads to excessive algal growth; when the algae die, the organic matter is decomposed by bacteria, a process which consumes a great deal of oxygen that could lead to a decrease in water quality and increases the threat to ecological life. Nitrite as an indicator of bacterial activity (Mahanta et al., 2014), ranged from 2.1 (coastal parts of Posm) to 13 (coastal parts of Ramin)  $\mu$ g/L in premonsoon, and from 4.4 to 9.6  $\mu$ g/L (coastal

parts of Konarak and entrance of Chabahar Bay) in post-monsoon, respectively (Figure 5).

Since few studies for nitrogen toxicity in environments of marine estuarine are available, the marine guideline is the same as the interim guideline proposed by CCME (2003). The 30-day average concentration of nitrate (as N), to protect freshwater aquatic life is 3.0 mg/L and its maximum concentration is 32.8 mg/L. The 30-day average concentration of nitrate (as N) to protect marine aquatic life is 3.7 mg/L. Minimum concentrations of nitrate in pre- $(2.2 \ \mu g/L)$  and post-monsoons  $(2.1 \ \mu g/L)$ were related to coastal parts of Tiss and deeper parts of Ramin, while its maximum values in pre- (25  $\mu$ g/L) and post-monsoons  $(15.3 \,\mu g/L)$  were detected at deeper parts of Ramin and entrance of Chabahar Bay, respectively (Figure 5).

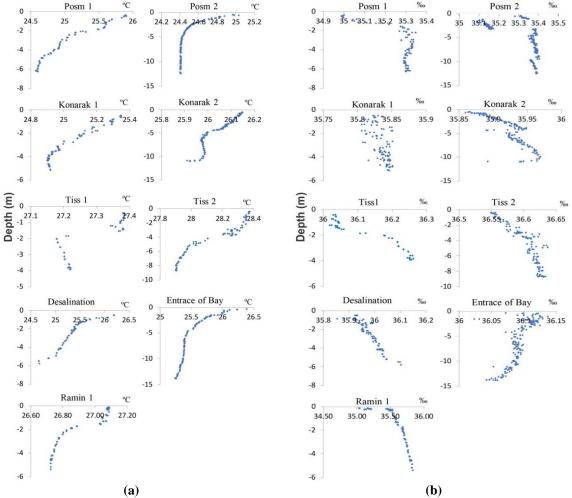


Fig. 3. Profiles of: a) Water temperature; b) Salinity; in pre Monsoon at the study area (May 2012)

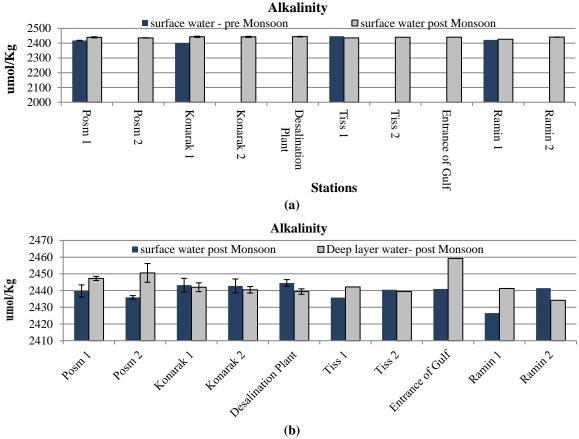
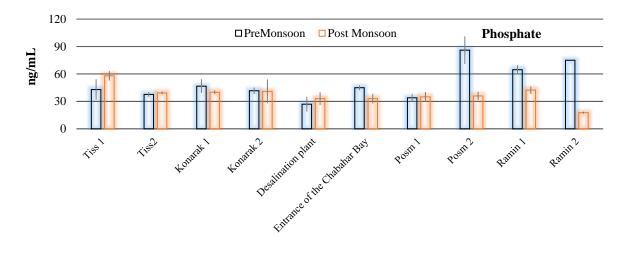
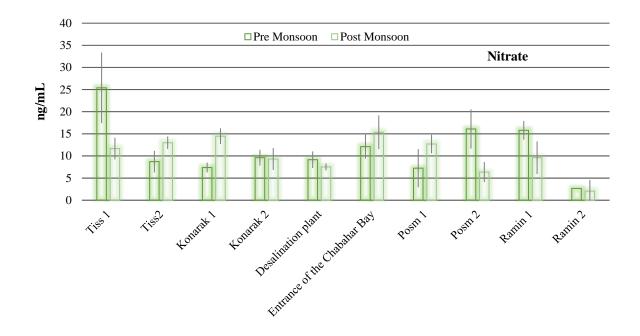


Fig. 4. Levels of Alkalinity: a) surface seawater in pre and post monsoon and; b) surface and near-bottom water in post monsoon (µmol/kg)

Nutrient concentration is highly variable in marine environment. In many cases, it shows a wide range of spatio-temporal variations depending on physical and biogeochemical processes. Based on Glodap2020 data, in the Gulf of Oman, NO<sub>3</sub> ranged from 0 at the surface to more than 35 umol/Kg (or 2100 ug/L at 1000 m), NO<sub>2</sub> ranged from 0 at surface to 0.75 umol/Kg (or 34 ug/L), PO<sub>4</sub> ranged from 0 at surface to 3 umol/Kg (or 285 ug/L at 1000 m), and Si ranged from 2 umol/Kg at surface to 150 umol/Kg at 3000 (56 to 4200 ug/L). Therefore, having values of near-zero for nutrients in surface samples is not unusual in this region; however, higher values of nutrients at the surface are expected during early winter in the northern Arabian Sea, due to winter convective mixing.





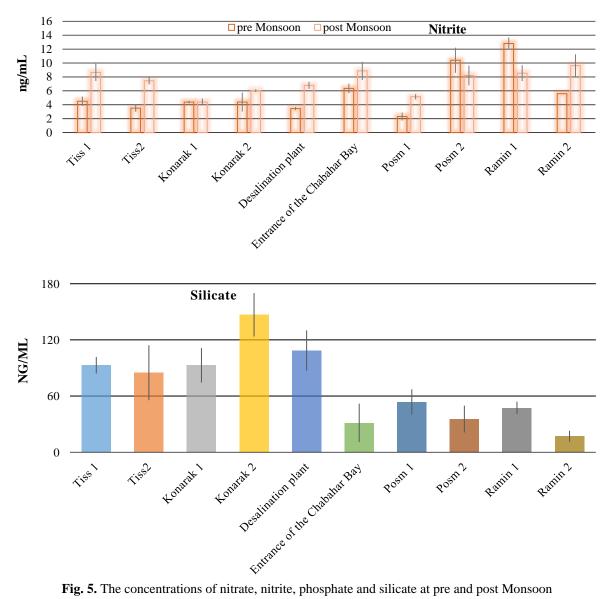


Fig. 5. The concentrations of nitrate, nitrite, phosphate and silicate at pre and post Monsoon

The concentrations of phosphate ranged from 27 (Desalination) to 86 (Posm), and from 17.7 (Ramin) to 58 (Tiss)  $\mu$ g/L in preand post-monsoons, respectively (Figure 5).

For Si, the mean value was significantly higher (P = 0.015) within the Bay, which could be the result of shallower environment inside the Bay. Probably, tidal currents could re-suspend bottom sediments in the shallow area, and release dissolved Si previously being trapped in sediments. The dilution effect in open sea stations could reduce this effect.

Average values of the nutrients in the present study were comparable to those reported in Gulf of Oman (nitrite: 2-26 and phosphate: 8-15  $\mu$ g/L) (Fazeli, 2010), and Persian Gulf at Strait of Hormoz (nitrite: 0.8-39.3, nitrate: 0-30 and phosphate: 0.5-55  $\mu$ g/L) (Sanjani, 2009).

The concentrations of chlorophyll-a and b in surface and bottom waters were determined at post-monsoon. The maximum chlorophyll-a in surface (0.64  $\mu$ g/L) and bottom water (1.20  $\mu$ g/L) were detected at Posm, while it was not detected at the stations inside Chabahar Bay. The maximum chlorophyll-b in surface (3.22  $\mu$ g/L) and bottom water (2.92  $\mu$ g/L) were related to deeper parts of Konarak (Figure 6).

Obtained results demonstrated that there significant correlations between were chlorophyll-a ( $r_2 = 1$ , p < 0.01) in surface and deep layer water, as well as chlorophyll-b ( $r_2 = 0.92$ , p < 0.01), but there were no correlations between chlorophyll-a and b in surface or deep layer waters (Table 3). It was revealed from statistical analysis that there were positive strong significant correlations between chlorophylls and dissolved oxygen content. Although pH variation within the studied area was 0.26, low pH variations inversely correlated with chlorophyll-a (p < 0.05) in surface water.

It might be necessary to conduct more investigations in a vast area during a longer period to be able to have an ecological justification on pH and chlorophyll variations. Chlorophyll-a was inversely correlated with temperature and positively correlated with salinity and dissolved oxygen (Table 3), which was in accordance with the results of Nurdin et al. (2013), who reported a negative correlation between temperature and chlorophylls.

Turbidity is an important operational the process affecting parameter of photosynthesis for algal growth (IEPA, 2001). Low levels of turbidity could be attributed to low wave actions and minimal turbulence, while high turbidity might be observed due to rainfall, riverine sediment loads, discharges of sewage and industrial waste or the presence of a large number of microorganisms (Olatayo, 2014). High turbidity reduces the light penetration, hence adversely influencing the primary, secondary and tertiary biological productions (Olatayo, 2014). Water clarity in pre- and post-monsoons were 0.9 (Tiss) -4.5 (Ramin) and 2.5 (Posm) -8.5 (Ramin) m, respectively. The clarity of water samples in pre- and post-monsoons varied from 0.9 (Tiss) to 4.5 (Ramin), and from 2.5 (Posm) to 8.5 (Ramin) m, respectively. Higher water clarity in post-monsoon could be related to its higher rain fall.

In this study, 66 phytoplankton genus and species belonging to 13 groups (Bacillarophyceae, Dinophyceae, Cyanophyceae, Alphaproteobacteria, Prymnesiophyceae, Zygnemophyceae and Chlorophyceae) were identified in premonsoon (Table 4).

The most dominant phytoplankton groups were Dinophyceae (73.30%) and Bacillariophyceae (22.30%). The maximum and minimum abundances of phytoplankton communities were detected at Tiss and Ramin stations, respectively. In postmonsoon, total phytoplankton abundance was  $1944333 \pm 5271$  (Cell L<sup>-1</sup>) and 994000 $\pm$  4475 (Cell L<sup>-1</sup>) in surface and depth (7 m) waters. In this survey, 66 species were identified; Bacillariophyceae (49.4%) and (44.5%) were the most Dinophyceae dominant phytoplankton groups, respectively.

In this survey, no significant correlation

(p > 0.05)was found between phytoplankton abundance, temperature and salinity (Table 5). The Mann-Whitney analysis showed that there was significant difference in phytoplankton density among different stations. Obtained results demonstrated that Tiss had higher distribution of phytoplankton compared to other sampling stations. Nutrient levels especially silicate and nitrate concentrations (Barcelos e Ramos et al., 2017; Mirzaei et al., 2017), originated from fishing and recreational beach as well as municipal sewers, might affect the higher phytoplankton abundance in this station.

### 4. Conclusions

In addition to receiving marine pollution from eastern and western neighborhoods (in pre- and post-monsoons), Chabahar Bay receives domestic, industrial, and in some cases, agricultural wastewater from its coastal areas, which can influence the quality of the water. In order to study the effect of anthropogenic influence on this ecosystem, in the present study, 10 stations were selected to assess the water quality. Results indicated that water salinity and pH (with narrow variation) were relatively higher in post-monsoon, especially in the stations inside the Chabahar Bay. Obtained results revealed that there was no significant change in pH values during two monitoring periods, pre- and postmonsoons. The pH values were within the range of 8.03 to 8.21, which was in accordance with Indian standards for sensitive ecological areas for coastal waters, such as Arabian Sea (Salvi et al., 2014). Appropriate pH and dissolved oxygen levels could express that the sampling area was suitable for marine life.

Temperature is one of the most important parameters for growth and survival of corals in marine water. Although temperature showed a temporal variation in accordance to the atmospheric variation and followed a seasonal trend, in the present study, the temperature was in appropriate range for corals as well as other marine diversity.

Moreover, results indicated that in premonsoon, minimum (0.03) and maximum (0.55) Shanoon indices for phytoplankton diversity were determined at Ramin and Tiss stations, respectively (Figure 7a). In post-monsoon, the minimum and maximum values of Shannon index were observed at Tiss and Konarak by 4.5 and 1.37, respectively (Figure 7b).

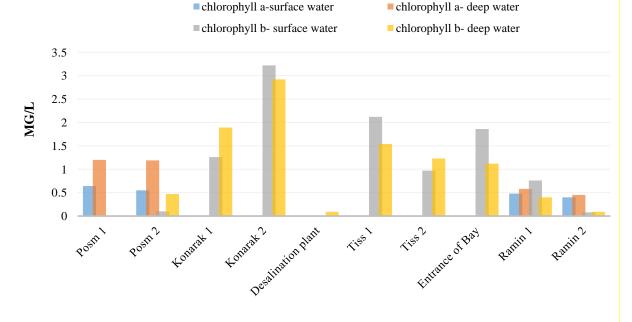


Fig. 6. Comparing chlorophyll a and b in surface and deep layer water at different stations in Post Monsoon

Chlorophylls and some environmental	Chloro	phyll-a	Chlorophyll-b			
factors	Surface water	Bottom water	Surface water	Bottom water		
Chlorophyll-a surface water	1.00					
Chlorophyll-a deep layer water	1.00**	1.00				
Chlorophyll-b surface water	-0.40	-0.40	1.00			
Chlorophyll-b bottom water	-0.20	-0.20	0.92**	1.00		
PH surface water	-0.74*	-0.74	-0.53	-0.51		
PH deep layer water	-0.32	-0.32	-0.56	-0.50		
Temperature surface water	-0.80**	-0.80**	-0.46	-0.67		
Temperature deep layer water	-1.00**	-1.00**	-0.50	-0.67		
Salinity surface water	0.95**	0.95**	0.03	0.25		
Salinity deep layer water	0.95**	0.95**	0.18	0.32		
Dissolved Oxygen surface water	1.00**	1.00**	-0.06	0.12		
Dissolved Oxygen deep layer water	0.80**	0.80**	-0.67	-0.40		

Table 3. Correlation between Chlorophylls and some environmental factors

\*: Significant at p < 0.05 and \*\*: P < 0.01 levels

\_

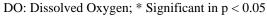
<b>Table 4.</b> The identified phytoplanktons

Class	Family	Genus	Pre (Abun dance± SE)	Post (Abun dance± SE)	Class	Family	Genus	Pre (Abun dance± SE)	Post (Abun dance± SE)
Dinophyc	Prorocentr	Prorocen	11000±	9083±2	Bacillariop	Achnanthac	Achnant	0	317±24
eae	aceae	trum	402	08	hyceae	eae	hes		
Dinophyc eae	Ceratiacea e	Ceratium spp.	3250±2 62	7833±4 10	Bacillariop hyceae	Catenulace ae	Amphor a	583±25	2733±6 0
Dinophyc	Protoperid	Protoper	9917±4	8950±2	Bacillariop	Melosirace			2717±2
eae	iniaceae	idinium	93	58	hyceae	ae	Melosira	0	26
Dinophyc	Dinophysi	Dinophy		783±	Bacillariop	Asterolamp	Asterola	_	
eae	aceae	sis spp.	167±9	23	hyceae	raceae	mpra	0	550±18
Dinophyc	Kolkwitzi	Preperid	385500	10850±	Bacillariop	Climacosph	Climaco	_	
eae	ellaceae	inium	±2360	311	hyceae	eniaceae	sphenia	0	67±4
Dinophyc	Gymnodi	Cochlodi			Bacillariop	Auriculace	-	_	
eae	niaceae	nium	0	133±13	hyceae	ae	Auricula	0	67±6
Dinophyc	Amphisol	Amphisol	0	100 0	Bacillariop	Fragilariace	<i>a</i> 1	0	4100±1
eae	eniaceae	enia	0	$133\pm8$	hyceae	ae	Synedra	0	19
Dinophyc	Noctiluca	Noctiluc	250 25	1467±3	Bacillariop	Thalassione	Thalassi	015 50	2633±2
eae	ceae	а	250±25	4	hyceae	mataceae	onema	917±59	46
Dinophyc	Calciodin	Scrippsie	0	5100±1	Bacillariop	Lauderiace		0	200 15
eae	ellaceae	lla	0	51	hyceae	ae	Lauderia	0	300±15
Dinophyc	Gymnodi	Gyrodini	167.16	10683±	Bacillariop	Thalassiosi	Thalasio	017.56	222.10
eae	niaceae	um	167±16	262	hyceae	raceae	sira	917±56	233±19
Dinophyc	Gonyaula	Gonyaul	0	(7) 1	Bacillariop	Biddulphia	Biddulph	0	92.6
eae	caceae	ax	0	67±4	hyceae	ceae	ia	0	83±6
Dinophyc	Gonyaula	Lingulod	$1000 \pm 7$	0	Bacillariop	Triceratiace	Lampris	0	150±15
eae	caceae	inium	9	0	hyceae	ae	cus	0	150±15
Dinophyc	Gonyaula	Alexandr	0	50±3	Bacillariop	Plagiotropi	Plagiotr	0	533±28
eae	caceae	ium	0	30±3	hyceae	daceae	opis	0	JJJ±20
Dinophyc	Dinotrich	Gymnodi	0	250±23	Bacillariop	Hemiaulace	Hemiaul	0	167±11
eae	aceae	nium	0	230±25	hyceae	ae	us	0	10/±11
Dinophyc	Gymnodi	Amphidi	0	$1100\pm$	Bacillariop	Pinnulariac	Pinnular	0	300±20
eae	niaceae	nium	0	70	hyceae	eae	ia	0	300±20
Dinophyc	Pyrophaca	Pyropha	1833±1	0	Bacillariop	Hemiaulace	Hemidis	0	150±13
eae	ceae	cus	33	0	hyceae	ae	cus	0	150±15
Bacillario	Naviculac	Navicula	$4000 \pm 2$	$6400\pm$	Bacillariop	Lithodesmi	Ditylum	2667±1	300±30
phyceae	eae	spp.	12	217	hyceae	aceae	Duytum	59	500±50
Bacillario	Rhizosole	Rhizosol	667±	1517±3	Bacillariop	Rhaphonei	Rhaphon	0	50±5
phyceae	niaceae	enia	47	2	hyceae	daceae	eis	U	50±5
Bacillario	Bacillaria	Pseudo-	917±62	3133±1	Bacillariop	Bacillariace	Cylindro	0	33±3
phyceae	ceae	nitzschia		03	hyceae	ae	theca	0	55-5
Bacillario	Fragilaria	Striatella	1750±9	17±1	Cyanophyc	Phormidiac	Trichode	0	283±18
phyceae	ceae		5		eae	eae	smium	0	205-10
Bacillario	Rhizosole	Guinardi	1750±4	4083±	Cyanophyc	Oscillatoria	Oscillato	750±75	500±32
phyceae	niaceae	а	7	165	eae	ceae	ria		

Bacillario	Bacillaria	Nitzschia	2000±7	13100±	Cyanophyc	Merismope	Aphanoc	1583±1	0
phyceae	ceae	111123011111	8	610	eae	diaceae	apsa	1363±1 58	0
Bacillario phyceae	Fragilaria ceae	Licmoph ora	0	283±26	Trebouxio phyceae	Chlorellace	Closteri opsis	0	4783±1 53
Bacillario	Coscinodi	Coscinod	7667±3	1517±4	Actinobact	Micrococcu	Microcr	0	$20033\pm$
phyceae	scaceae	oscus	35	2	eria	ceae	ocis	0	767
Bacillario phyceae	Coscinodi scaceae	Lennoxia	333±23	0	Alphaprote obacteria	Sphingomo nadaceae	Rhizomo nas	500±50	0
Bacillario phyceae	Chaetocer otaceae	Chaetoce ros spp.	0	5983±1 87	Chrysophy ceae	Dinobryace ae	Dinobry on	0	300±8
Bacillario phyceae	Leptocyli ndraceae	Leptocyli ndrus	0	3150± 83	Florideoph vceae	Lemaneace ae	Lemanea	0	250±16
Bacillario phyceae	Amphiole uraceae	Amphipr ora	0	833±55	Xanthophy ceae	Heteropedi aceae	Heteroc occus	0	250±16
Bacillario	Skeletone maceae	Skeleton ema	0	1367±1 09	Zygnemop hyceae	Closteriace	Closteri um	250±25	0
Bacillario phyceae	Hemiaula ceae	Eucampi a	167±96	467± 30	Zygnemop hyceae	Desmidiace ae	Cosmari um	417±41	0
Bacillario phyceae	Diploneid aceae	Diplonei s	0	567± 19	Chlorophy ceae	Radiococca ceae	Coenoco ccus	2167±2 16	0
Bacillario	Naviculac	Pleurosi	417.21	$4067 \pm$	Prymnesio	Rhabdosph	Coronos	1917±1	0
phyceae	eae	gma	417±31	213	phyceae	aeraceae	phaera	91	0
Bacillario	Hemiaula	Cerataul	0	750±38	Dictyocho	Dictyochac	Dichtyoc	0	1033±2
phyceae	ceae	ina	0	750±58	phyceae	eae	ha	0	9

 Table 5. Correlation coefficients between phytoplankton abundance and physic- chemical parameters in pre and post Monsoons

	Pre	Monsoon					
Parameter	Phytoplankton	Temperature	pН	Phytoplankton	Temperature	pН	Salinity
Temperature	0.8			0.65			
pН	0	0.8		0.4	0.03		
Salinity	0.05	0.68	0.05	0.59		0.16	
DO	-	-	-	0.50	0.86	0.19	0.01 *



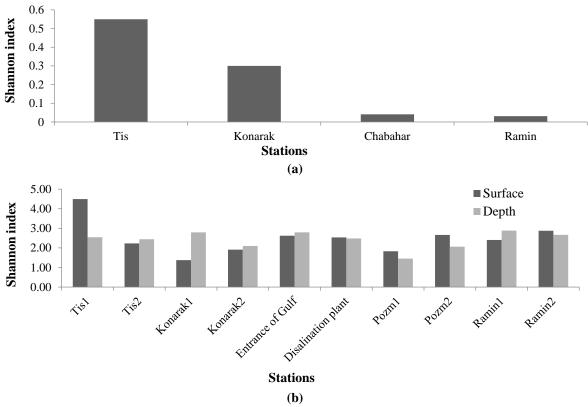


Fig. 7. Shannon index values in the sampling stations in: a) Pre Monsoon and; b) Post Monsoon

The higher levels of nutrients and subsequently phytoplankton abundance in surface water in post-monsoon could be due to strong wind currents, which mix surface and deep layer waters.

Excess phosphate level can accelerate the growth rate of algae and can result in death and decay of marine benthos and fishes, and might reduce water quality since it decreases the levels of dissolved oxygen. Phosphorous level higher than 1 mg/L can cause excessive plant growth and eutrophication (Ron Fleming, 1999). The average phosphorous level in the present study (0.09  $\mu$ g/L) was much less than defined values in guidelines.

Low variations of water parameters, and low dissolved organic carbon (< 2 mg/L) have provided appropriate conditions to establish desalination plant in this area. In general, the physicochemical parameters of water samples from all stations indicated that water quality falls within the stipulated range of acceptability and sampling area can be classified as a good, stable and healthy aquatic ecosystem.

There should be a constant monitoring of the physicochemical parameters of the studied area in the future, due to the increase in anthropogenic activities around this area. For further investigations, it is recommended to assess the offshore physical, chemical, and biological parameters (> 50 m depth) for cage fish farming.

#### 5. Acknowledgement

The authors gratefully acknowledge financial support from INIOAS-Tehran. In addition, the authors wish to thank all the colleagues who have contributed to in-situ processes.

#### 6. References

Abowei, J. (2010). "Salinity, dissolved oxygen, pH and surface water temperature conditions in Nkoro River, Niger Delta, Nigeria", *Advance Journal of Food Science and Technology*, 2(1), 36-40.

- Agah, H. (2018). "Assessment of trace metals accumulation in water and sediments of Chabahar Bay, Makoran", *Indian Journal of Geo Marine Science*, 47(11), 2230-2236.
- Agah, H., Rahmanpour S. and Sheijooni Fumani, N. (2014). "Organic carbon and organic matter contents in sediments of the Strait of Hormoz, the Persian Gulf", *Journal of the Persian Gulf*, 4(13), 31-37.
- AlgaBbase, https://www.algaebase.org/
- Al-Kandari, M., Al-Yamani, F. and Al-Rifaie, K. (2009). *Marine phytoplankton atlas of Kuwait's waters*, Kuwait Institute for Scientific Research, Kuwait, ISBN 99906-41-24-2.
- APHA (American Public Health Association).
   (1995), Standard methods for the examination of water and waste water, 19<sup>th</sup> Editions, Washington, DC.
- Barcelos e Ramos, J., Schulz, K.G., Voss, M., Narciso, Á., Müller, M.N., Reis, F.V., Cachão, M. and Azevedo, E.B. (2017). "Nutrient-specific responses of a phytoplankton community: A case study of the North Atlantic Gyre, Azores" *Journal of Plankton Research*, 39(4), 744-761.
- Basu, S. and Mackey, K.R.M. (2018). "Phytoplankton as key mediators of the biological carbon pump: their responses to a climate changing", *Sustainability*, 10(3), 869.
- Burt, J.A., Coles, S., Lavieren, H., Taylor, O., Looker E. and Samimi-Namin, K. (2016).
  "Oman's coral reefs: A unique ecosystem challenged by natural and man-related stresses and in need of conservation", *Marine Pollution Bulletin*, 105(2), 498-506.
- CCME (Canadian Council of Ministers of the Environment). (2003). Canadian water quality guideline for the protection of aquatic life, Winnipeg, MB.
- EPA, (1970). Conference in the Matter of Pollution of Lake Superior and Its Tributary basin Minnesota, Wisconisn Michigan, 155, 29-30 April.
- Dickson, A.G., Sabin, C.L. and Christian, J.R. (2007). Guide to best practices for ocean CO<sub>2</sub> measurements, PICES Special Publication, 191.
- Ding, H. and Elmore, A.J. (2015). "Spatio-temporal patterns in water surface temperature from Landsat time series data in the Chesapeake Bay, USA", *Remote Sensing of Environment*, 168, 335-348.
- Efe, S.I., Ogban, F.E., Horsfall, M.J. and Akporhonor, E.E. (2005). "Seasonal variations of physicchemical characteristics in water resources quality in Western Niger Delta Region, Nigeria", *Journal of Applied Science and Environment*, 9(1), 191-195.
- Emara, H.I. (2010). "Nutrient salts, inorganic and organic carbon contents in the waters of the Persian Gulf and the Gulf of Oman", *Journal of the Persian Gulf (Marine Science)*, 1(2), 33-44.

- Fazeli, N., Marnani, H.R. and Sanjani, S. (2010). "Seasonal variation of copepoda in Chabahar Bay-Gulf of Oman", *Jordan Journal of Biological Sciences*, 147(616), 1-24.
- Fazeli, N., (2008). "Ecological studies of zooplanktons in Chabahar Bay", M.Sc. Thesis, University of Marine Science, Khorramshahr, Iran.
- Friedrich, J., Janssen, F., Aleynik, D., Bange, H.W., Boltacheva, N., Çagatay, M.N., Dale, A.W., Etiope, G., Erdem, Z., Geraga, M., Gilli, A., Gomoiu, M.T., Hall, P.O.J., Hansson, D., Holtappels, Y.H.M.M., Kirf, K., Kononets, M., Konovalov, S., Lichtschlag, A., Livingstone, D.M., Marinaro, G., Mazlumyan, S., Naeher, S., North, R.P., Papatheodorou, G., Pfannkuche, O., Prien, R., Rehder, G., Schubert, C.J., Soltwede, T., Sommer, S., Stahl, H., Stanev, E.V., Teaca, A., Tengberg, A., Waldmann, C., Wehrli, B. and Wenzhöfer, F. (2014). "Investigating hypoxia in aquatic environments: Diverse approaches to addressing complex phenomenon", а Biogeosciences, 11(4), 1215-1259.
- IEPA (Ireland Environmental Protection Agency). (2001). *Parameters of water quality: Interpretation and standards*, Environmental Protection Agency Johnstown, pp. 133.
- Iqbal, F., Ali, M., Salam, A., Khan, B.A., Ahmad, S. and Qamar, M. (2004). "Seasonal variations of physico-chemical characteristics of river Soan water at Dhoak Pathan bridge (Chakwal), Pakistan", *International Journal of Agriculture* and Biology, 6(1), 89-92.
- Karlson, B., Cusack, C. and Bresnan, E. (2010). "Microscopic and molecular methods for quantitative phytoplankton analysis", UNESCO, IOC Manuals and Guides, No. 55. (IOC/2010/MG/55)
- Khan, M.N. and Mohammad, F. (2014), *Eutrophication: Challenges and solutions*, Chapter 1, A.A. Ansari and S.S. Gill (eds.), Eutrophication: Causes, Consequences and Control, pp. 1-15.
- Mahanta, D., Jena, J.K. and Das, B.K. (2014). "Evaluation and efficacy study of Nitrifying Bacteria in freshwater Aquaculture system", *International Journal of Current Microbiology* and Applied Science, 3(7), 962-969.
- Mearns, A.J., Reish, D.J., Oshida, P.S., Ginn, T., Rempel-Hester, M.A., Arthur, C. and Rutherford, N. (2013). "Effects of pollution on marine organisms", *Water Environment Research*, 85(10), 1828-1933.
- Mirzaei, M.R., Seraji, F., Erfanifar, E., Aminirad, T., Aminikhoei, Z. and Azhang, B. (2017), "Assessing phytoplankton community structure in relation to hydrographic parameters and seasonal variation (Pre and Post Monsoon)", *Biodiversitas*, 18(2), 507-513.
- NOAA, Coral Reef Watch, (2019). Monthly and

annual composites of 5 km satellite coral bleaching heat stress products (Version 3.1), 1985 to 2019, Data set accessed 2019-19-10 at https://coralreefwatch.noaa.gov/satellite/hdf/ind ex.php.

- Nowrouzi, S. and Valavi, H. (2011). "Effects of environmental factors on phytoplankton abundance and diversity in Kaftar Lake", *Journal of Fisheries and Aquatic Sciences*, 6, 130-140.
- Nurdin, S., Mustapha, M.A. and Lihan, T. (2013). "Spatial and temporal variability of sea surface temperature in Makassar Strait, Indonesia", In Proceedings of 12<sup>th</sup> International University Malaysia Terengganu Annual Symposium, UMTAS 2013 Advancements in Marines and Freshwater Sciences, pp. 26-28.
- Olatayo, A.A. (2014). "Assessment of physicochemical parameters of waters in ilaje local government area of Ondo State, Nigeria", *International Journal of Fisheries and Aquatic Studies*, 1(5), 84-92.
- Olsen, A., Lange, N., Key, R.M., Tanhua, T.H., Bittig, C., Kozyr, A., Álvarez, M. Azetsu-Scott, K., Becker, S., Brown, P.J., Carter, B.R., Cotrim da Cunha, L., Feely, R.A., van Heuven, S., Hoppema, M., Ishii, M., Jeansson, E., Jutterström, S., Landa, C.S., Lauvset, S.K., Michaelis, P., Murata, A., Pérez, F.F., Pfeil, B., Schirnick, C., Steinfeldt, R., Suzuki, T., Tilbrook, B., Velo, A., Wanninkhof, R. and Woosley, R.J. (2020). "An updated version of the global interior ocean biogeochemical data product, GLODAPv2.2020", *Earth System Science Data*, 12(4), 3653-3678.
- Omer, W.M.M. (2010). "Ocean acidification in the Arabian Sea and the Red Sea", M.Sc. Thesis, Faculty of Mathematics and Natural Sciences, Geophysical Institute, Chemical Oceanography. University of Beregen, Norway.
- Raven, J., Caldeira, K., Elder\_eld, H., Hoegh-Guldberg, O., Liss, P., Riebesell, U., Shepherd, J., Turley, C., Watson, A., Heap, R., Banes, R. and Quinn, R. (2005). Ocean acidification due to increasing atmospheric carbon dioxide, Technical Report, The Royal Society, Working Group, United Kingdom.
- ROPME (Regional Organization for the Protection of the Marine Environment). (1999). Manual of oceanographic observations and pollutant analyses methods (MOOPAM), Kuwait: Regional Organization for the Protection of the Marine Environment, 3<sup>rd</sup> Edition.
- ROPME (Regional Organization for the Protection of the Marine Environment). (2012). "Spatial distribution of Chlorophyll-a in the ROPME Sea Area", Technical Report: No. 4, ROPME/GC-14/7.
- Saleh, A., Vajed Samiei, J., Amini-Yekta, F., Seyed Hashtroudi, M., Chen, C.-T.A. and Fumani, N.S.

(2020). "The carbonate system on the coral patches and rocky intertidal habitats of the northern Persian Gulf: Implications for ocean acidification studies", *Marine Pollution Bulletin*, 151, 110834.

- Salvi, H., Patel, R., Thakur, B., Shah, K. and Parmar, D. (2014). "Assessment of coastal water quality parameters of selected areas of marine national park and sanctuary (Okha, Sikka and Khijadiya)", SSRN 2456093 Electronic Journal.
- Sanjani, M., (2009). Report of oceanic patrol of maritime organizations around Islands near strait of Hormoz (Persian Gulf), Iranian National Institute for Oceanography, Tehran, Iran.
- Shanthi, R., Vinoth, Gowtham, A., Senthilnathan, L., Poornima1, D., Sarangi, R.K. and Thangaradjou, T. (2014). "Experimental studies on silicate as the limiting nutrient in altering phytoplankton community structure", *International Journal of Advanced Earth Science* and Engineering, 3(1), 225-239.
- Xue, L. and Cai, W.J. (2020). "Total alkalinity minus dissolved inorganic carbon as a proxy for deciphering ocean acidification mechanisms", *Marine Chemistry*, 222, 103791.



This article is an open-access article distributed under the terms and conditions of the Creative Commons Attribution (CC-BY) license.